

*Biochimica et Biophysica Acta*, 484 (1977) 417–422  
 © Elsevier/North-Holland Biomedical Press

BBA 68238

## SYNTHETIC INHIBITORS OF TRYPSIN, PLASMIN, KALLIKREIN, THROMBIN, $C_1\bar{r}$ , AND $C_1$ ESTERASE

YOSHIYUKI TAMURA <sup>a</sup>, MASAYUKI HIRADO <sup>a</sup>, KAZUKO OKAMURA <sup>a</sup>, YOSHIHIRO MINATO <sup>a</sup> and SETSURO FUJII <sup>b</sup>

<sup>a</sup> Department of Enzyme Physiology, Institute for Enzyme Research, School of Medicine, Tokushima University, Tokushima, and <sup>b</sup> Institute for Protein Research, Osaka University, Osaka (Japan)

(Received September 21st, 1976)

(Revised manuscript received May 10th, 1977)

### Summary

*p*-Carbethoxyphenyl  $\epsilon$ -guanidinocaproate and *p*-(*p*'-guanidinobenzoyloxy)-phenyl derivatives were prepared, and their inhibitory effects on trypsin, plasmin, plasma kallikrein, thrombin,  $C_1\bar{r}$  and  $C_1$  esterase were examined. Among the various inhibitors tested, *p*-nitrophenyl *p*'-guanidinobenzoate, *N,N*-dimethylamino *p*-(*p*'-guanidinobenzoyloxy)-benzoyl glycolate and *N,N*-dimethylamino *p*-(*p*'-guanidinobenzoyloxy)-benzilcarbonyloxy glycolate were the most effective inhibitors of trypsin, plasmin, plasma kallikrein and thrombin, and they strongly inhibited the esterolytic activities of  $C_1\bar{r}$  and  $C_1$  esterase.

### Introduction

Previously [1,2], we reported the strong, reversible inhibitory effects of various  $\omega$ -guanidino acid esters on trypsin, plasmin, plasma kallikrein and thrombin. Among the various esters examined, aromatic esters, such as *p*-carbethoxyphenyl  $\epsilon$ -guanidinocaproate monophosphate, were more inhibitory than aliphatic esters. Moreover, aromatic esters of  $\epsilon$ -guanidinocaproic acid and *trans*-4-aminomethylcyclohexane-carboxylic acid (AMCHA) caused strong, competitive inhibition of  $C_1$  esterase and the conversion of  $C_1$  to  $C_1$  esterase and the strongest inhibitor was the phenyl ester of  $\epsilon$ -guanidinocaproic acid [3].

Mares-Guia and Shaw [4] reported the irreversible inhibitory effect of ethyl

---

Abbreviations: TosArgOMe,  $N^\alpha$ -tosyl-L-arginine methyl ester; N-AcTyrOEt,  $N^\alpha$ -acetyl-L-tyrosine ethyl ester; N-AcArgOMe,  $N^\alpha$ -acetyl-L-arginine methyl ester; AMCHA, *trans*-4-aminomethylcyclohexane-carboxylic acid;  $\epsilon$ -GCA-CEP, *p*-carbethoxyphenyl  $\epsilon$ -guanidinocaproate; *p*'-GB-NP, *p*-nitrophenyl *p*'-guanidinobenzoate; *p*'-GB-DBoG, *N,N*-dimethylamino *p*-(*p*'-guanidinobenzoyloxy)-benzoyl glycolate; *p*'-GB-DBiG, *N,N*-dimethylamino *p*-(*p*'-guanidinobenzoyloxy)-benzilcarbonyloxy glycolate.

*p*-guanidinobenzoate on trypsin, and Chase and Shaw [5] reported that of *p*-nitrophenyl-*p*'-guanidinobenzoate. Bing [6] reported that a highly purified subunit of activated human complement, component C<sub>1</sub> (C<sub>1</sub> $\bar{s}$ ), hydrolyzed *p*-nitrophenyl N $\alpha$ -carbobenzoxy-L-tyrosinate, and that the reaction was inhibited competitively by guanidine, amidine and a variety of aromatic compounds of low molecular weight.

This paper reports that *p*-(*p*'-guanidinobenzoyloxy)-phenyl derivatives are stronger inhibitors of trypsin, plasmin, plasma kallikrein and thrombin than *p*-carbethoxyphenyl  $\epsilon$ -guanidinocaproate monophosphate, and that they inhibit C<sub>1</sub> esterase as much as the phenyl ester of  $\epsilon$ -guanidinocaproic acid. Studies on the inhibitory effects of these compounds on C<sub>1</sub> $\bar{r}$ , a subcomponent of the first component of human complement are also described.

## Materials and Methods

### Enzymes

Human plasmin [7], plasma kallikrein [7], C<sub>1</sub> esterase [8] and C<sub>1</sub> $\bar{r}$  [9] were prepared as described previously. Bovine trypsin (type I) was purchased from Sigma Chemical Co., and dissolved in 0.1 M sodium phosphate buffer, pH 7.4. Human thrombin was purchased from the Green Cross Corporation, Osaka, Japan and dissolved in 0.1 M sodium phosphate buffer, pH 7.4.

### Substrates and inhibitors

N $\alpha$ -Tosyl-L-arginine methyl ester (TosArgOMe), N $\alpha$ -acetyl-L-tyrosine ethyl ester (N-AcTyrOEt), and N $\alpha$ -acetyl-L-arginine methyl ester (N-AcArgOMe) were purchased from the Foundation for Promotion of Protein Research, Institute for Protein Research, Osaka University, Osaka, Japan. Casein was purchased from E. Merck Japan Ltd, and bovine fibrinogen (fraction I) from Sigma Chemical Co. *p*-Carbethoxyphenyl  $\epsilon$ -guanidinocaproate ( $\epsilon$ -GCA-CEP), *p*-nitrophenyl *p*'-guanidinobenzoate (*p*'-GB-NP), *N,N*-dimethylamino *p*-(*p*'-guanidinobenzoyloxy)-benzoyl glycolate (*p*'-GB-DBoG) and *N,N*-dimethylamino *p*-(*p*'-guani-

Compound	Structure
$\epsilon$ -GCA-CEP	$\begin{array}{c} \text{HN} \\ \parallel \\ \text{C} - \text{NH} - \text{CH}_2 - \text{CH}_2 - \text{CH}_2 - \text{CH}_2 - \text{CH}_2 - \text{COO} - \text{C}_6\text{H}_4 - \text{COO} - \text{Et} \\ \diagup \\ \text{H}_2\text{N} \end{array}$
<i>p</i> '-GB-NP	$\begin{array}{c} \text{HN} \\ \parallel \\ \text{C} - \text{NH} - \text{C}_6\text{H}_4 - \text{COO} - \text{C}_6\text{H}_4 - \text{NO}_2 \\ \diagup \\ \text{H}_2\text{N} \end{array}$
<i>p</i> '-GB-DBoG	$\begin{array}{c} \text{HN} \\ \parallel \\ \text{C} - \text{NH} - \text{C}_6\text{H}_4 - \text{COO} - \text{C}_6\text{H}_4 - \text{COO} - \text{CH}_2 - \text{CO} - \text{N}(\text{CH}_3)_2 \\ \diagup \\ \text{H}_2\text{N} \end{array}$
<i>p</i> '-GB-DBiG	$\begin{array}{c} \text{HN} \\ \parallel \\ \text{C} - \text{NH} - \text{C}_6\text{H}_4 - \text{COO} - \text{C}_6\text{H}_4 - \text{CH}_2 - \text{COO} - \text{CH}_2 - \text{CO} - \text{N}(\text{CH}_3)_2 \\ \diagup \\ \text{H}_2\text{N} \end{array}$

Fig. 1. Structural formulae of  $\epsilon$ -GCA-CEP, *p*'-GB-NP, *p*'-GB-DBoG, and *p*'-GB-DBiG.

dinobenzoyloxy)-benzilcarbonyloxy glycolate (*p'*-GB-DBiG) were prepared in the Research Laboratories of Ono Pharmaceutical Company, Osaka, Japan. The structural formulas of these compounds are shown in Fig. 1.

#### Experiments on inhibition of various enzymes

The rates of hydrolysis of TosArgOMe by trypsin, plasmin, plasma kallikrein and thrombin were determined as described previously [10], at a substrate concentration of 10 mM; The rates of hydrolysis of *N*-AcArgOMe by  $C_1\bar{r}$  and *N*-AcTyrOEt by  $C_1$  esterase were determined as described previously [9], at a substrate concentration of 10 mM. The caseinolysis of trypsin and fibrinogenolysis of plasmin were determined as described previously [7]. The final concentrations of both casein and fibrinogen were 2%. To measure inhibitor, mixtures of enzyme solution and inhibitor were preincubated at 37°C for 5 min and then residual enzyme activities were determined, as described above.

The  $K_m$  values were determined from the Lineweaver-Burk [11] plot of the results. The  $K_i$  values were determined as described by Dixon [12].

#### Results and Discussion

The effects of  $\epsilon$ -GCA-CEP and *p*-(*p'*-guanidinobenzoyloxy)-phenyl derivatives on the esterolysis of various enzymes were examined, and the concentrations required for 50% inhibition are shown in Table I. The concentrations of *p'*-GB-DBiG required for 50% inhibition of the caseinolysis of trypsin and fibrinogenolysis of plasmin were  $3.0 \cdot 10^{-8}$  M and  $2.5 \cdot 10^{-7}$  M, respectively. This indicated that the intensity of the inhibitor activity on esterolysis and proteolysis was almost the same.

Previously [2], we examined the inhibitory effects of various esters of  $\omega$ -guanidino acids on trypsin, plasmin, plasma kallikrein and thrombin and found that among the various inhibitors tested, phenyl and *p*-carbethoxyphenyl  $\epsilon$ -guanidinocaproate were the most inhibitory to these four enzymes.

On the other hand, *p*-(*p'*-guanidinobenzoyloxy)-phenyl derivatives inhibited these four enzymes more than  $\epsilon$ -GCA-CEP, as shown in Table I. *p'*-GB-DBoG strongly inhibited trypsin, plasmin and thrombin: the concentrations causing 50% inhibition of TosArgOMe hydrolysis by trypsin, plasmin and thrombin were  $1.95 \cdot 10^{-9}$ ,  $3.6 \cdot 10^{-8}$ ,  $1.2 \cdot 10^{-6}$  M, respectively. The concentration of

TABLE I

#### CONCENTRATIONS OF INHIBITORS FOR 50% INHIBITION OF ESTEROLYSIS

Incubations were carried out in 0.1 M sodium phosphate buffer, pH 7.4, at 37°C. TosArgOMe (10 mM) was used as substrate for trypsin, plasmin, kallikrein, and thrombin, *N*-AcArgOMe (10 mM) for  $C_1\bar{r}$ ; and *N*-AcTyrOEt (10 mM) for  $C_1$  esterase.

Inhibitor	Inhibitor concentration for 50% inhibition (M)					
	Trypsin	Plasmin	Plasma kallikrein	Thrombin	$C_1\bar{r}$	$C_1$ esterase
$\epsilon$ -GCA-CEP	$9.4 \cdot 10^{-6}$	$3.0 \cdot 10^{-5}$	$4.1 \cdot 10^{-5}$	$1.1 \cdot 10^{-4}$		$5.2 \cdot 10^{-4}$
<i>p'</i> -GB-NP	$2.7 \cdot 10^{-8}$	$4.3 \cdot 10^{-8}$	$3.8 \cdot 10^{-7}$	$2.4 \cdot 10^{-6}$		
<i>p'</i> -GB-DBoG	$1.95 \cdot 10^{-9}$	$3.6 \cdot 10^{-8}$		$1.2 \cdot 10^{-6}$		$3.0 \cdot 10^{-5}$
<i>p'</i> -GB-DBiG	$3.2 \cdot 10^{-8}$	$1.9 \cdot 10^{-7}$	$3.2 \cdot 10^{-7}$	$5.0 \cdot 10^{-5}$	$4.4 \cdot 10^{-6}$	$3.4 \cdot 10^{-5}$

TABLE II

 $K_i$  VALUES FOR TRYPSIN, PLASMIN,  $C_{1r}$  AND  $C_1$  ESTERASE

$K_i$  values were estimated in 0.1 M sodium phosphate buffer, pH 7.4, at 37°C. TosArgOMe was used as substrate for trypsin and plasmin, *N*-AcArgOMe for  $C_{1r}$ , and *N*-AcTyrOEt for  $C_1$  esterase.

Inhibitor	$K_i$ (M)			
	Trypsin	Plasmin	$C_{1r}$	$C_1$ esterase
$\epsilon$ -GCA-CEP	$2.6 \cdot 10^{-6}$			
<i>p'</i> -GB-DBoG	$8.1 \cdot 10^{-7}$			
<i>p'</i> -GB-DBiG	$3.3 \cdot 10^{-8}$	$3.8 \cdot 10^{-8}$	$5.0 \cdot 10^{-8}$	$1.2 \cdot 10^{-4}$

*p'*-GB-DBiG for 50% inhibition of TosArgOMe hydrolysis by plasma kallikrein was  $3.2 \cdot 10^{-7}$  M. On the other hand, the concentrations of  $\epsilon$ -GCA-CEP for 50% inhibition of TosArgOMe hydrolysis by trypsin, plasmin, plasma kallikrein and thrombin were  $9.4 \cdot 10^{-6}$ ,  $3.0 \cdot 10^{-5}$ ,  $4.1 \cdot 10^{-5}$  and  $1.1 \cdot 10^{-4}$  M, respectively, as shown in Table I.

Previously, we reported [3] that aromatic esters of  $\epsilon$ -guanidinocaproic acid and *trans*-4-aminomethylcyclohexane carboxylic acid (AMCHA) caused strong, competitive inhibition of  $C_1$  esterase and the conversion of  $C_1$  to  $C_{1r}$  esterase. The phenyl ester of  $\epsilon$ -guanidinocaproic acid was the strongest inhibitor and the concentration causing 50% inhibition of *N*-AcTyrOEt hydrolysis of  $C_1$  esterase was approximately  $1 \cdot 10^{-4}$  M. On the other hand, as shown in Table I, *p'*-GB-DBoG and *p'*-GB-DBiG inhibited  $C_1$  esterase more than phenyl  $\epsilon$ -guanidinocaproate and  $\epsilon$ -GCA-CEP: the concentrations of *p'*-GB-DBoG and *p'*-GB-DBiG for 50% inhibition of  $C_1$  esterase were  $3.0 \cdot 10^{-5}$  and  $3.4 \cdot 10^{-5}$  M, respectively.

Furthermore, *p'*-GB-DBiG inhibited  $C_{1r}$  more than  $C_1$  esterase: the concentrations causing 50% inhibition of *N*-AcArgOMe hydrolysis by  $C_{1r}$  and *N*-AcTyrOEt hydrolysis by  $C_1$  esterase were  $4.4 \cdot 10^{-6}$  and  $3.4 \cdot 10^{-5}$  M, respectively, as shown in Table I.

$K_i$  values of these inhibitors on esterolysis using various substrates were then determined. Table II shows that the  $K_i$  value of  $\epsilon$ -GCA-CEP for trypsin hydrolytic activity was  $2.6 \cdot 10^{-6}$  M, the  $K_i$  value of *p'*-GB-DBoG was  $8.1 \cdot 10^{-7}$  M, and  $K_i$  value of *p'*-GB-DBiG was  $3.3 \cdot 10^{-8}$  M.  $K_i$  values of *p'*-GB-DBiG for plasmin,  $C_{1r}$ , and  $C_1$  esterase were  $3.8 \cdot 10^{-8}$ ,  $5.0 \cdot 10^{-8}$ , and  $1.2 \cdot 10^{-4}$  M, respectively. Modes of inhibitions by these compounds were all competitive.  $K_m$  values for trypsin, plasmin,  $C_{1r}$  and  $C_1$  esterase were estimated as shown in

TABLE III

 $K_m$  VALUES FOR TRYPSIN, PLASMIN,  $C_{1r}$  AND  $C_1$  ESTERASE

$K_m$  values were estimated in 0.1 M sodium phosphate buffer, pH 7.4, at 37°C.

Enzyme	Substrate	$K_m$ (M)
Trypsin	TosArgOMe	$3.1 \cdot 10^{-3}$
Plasmin	TosArgOMe	$6.0 \cdot 10^{-3}$
$C_{1r}$	<i>N</i> -AcArgOMe	$1.2 \cdot 10^{-2}$
$C_1$ esterase	<i>N</i> -AcTyrOEt	$4.2 \cdot 10^{-2}$

TABLE IV

EFFECT OF DIALYSIS ON ESTEROLYSIS OF  $\epsilon$ -GCA-CEP AND  $p'$ -GB-DBoG TREATED TRYPSIN

A mixture of trypsin (20  $\mu\text{g/ml}$ ) and inhibitor ( $10^{-3}$  M) in 0.1 M borate buffer containing 0.001 M  $\text{CaCl}_2$ , pH 8.5 was incubated at  $37^\circ\text{C}$  for 5 min and then dialyzed against the same buffer overnight, at  $4^\circ\text{C}$ . A mixture of 0.1 ml of dialyzate and 10 mM TosArgOMe were incubated at  $37^\circ\text{C}$  for 30 min and then the esterase activities of trypsin were determined. Values are shown in  $\mu\text{mol}$  per 0.1 ml.

	Buffer *	Inhibitor ( $10^{-3}$ M)	
		$\epsilon$ -GCA-CEP	$p'$ -GB-DBoG
Dialysis (—)	19.94	3.62	2.36
Dialysis (+)	19.90	19.80	2.45

\* 0.1 M borate buffer containing 0.001 M  $\text{CaCl}_2$ , pH 8.5

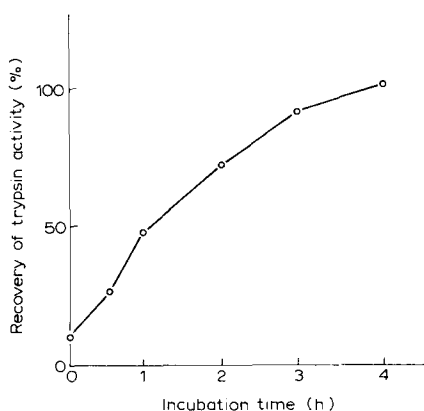


Fig. 2. Effect of incubation on esterolysis of the inhibitor · trypsin complex. A mixture of trypsin (20  $\mu\text{g/ml}$ ) and  $p'$ -GB-DBoG ( $10^{-3}$  M) in 0.1 M borate buffer containing 0.001 M  $\text{CaCl}_2$ , pH 8.5 was incubated at  $37^\circ\text{C}$  for 5 min and then dialyzed against the same buffer overnight, at  $4^\circ\text{C}$ . Amounts of 0.1 ml of dialyzate were incubated at  $37^\circ\text{C}$  for the indicated time intervals and esterase activities of trypsin were determined following incubation of the mixture at  $37^\circ\text{C}$  for 30 min after addition of 10 mM TosArgOMe.

Table III. The  $K_m$  value for trypsin with TosArgOMe as substrate was  $3.1 \cdot 10^{-3}$ ,  $6.0 \cdot 10^{-3}$  M for plasmin with TosArgOMe,  $1.2 \cdot 10^{-2}$  M for  $\text{C}_1\text{r}$  with  $N$ -AcArg-OMe, and  $4.2 \cdot 10^{-2}$  M for  $\text{C}_1$  esterase with  $N$ -AcTyrOEt.

The various compounds described above were reversible inhibitors of trypsin, plasmin, plasma kallikrein, thrombin,  $\text{C}_1\text{r}$  and  $\text{C}_1$  esterase. For example, as shown in Table IV,  $\epsilon$ -GCA-CEP was easily removed from the inhibited enzyme by dialysis.  $p'$ -GB-DBoG was not removed from the inhibited enzyme by dialysis, but after dialysis it was completely removed by incubating the inhibited enzyme at  $37^\circ\text{C}$  for 4 h, as shown in Fig. 2.

Experiments are now in progress on the mechanisms of inhibition of the various enzymes and the behaviors of these inhibitors.

## References

- 1 Muramatsu, M. and Fujii, S. (1971) Biochem. Biophys. Acta 242, 203–208
- 2 Muramatsu, M. and Fujii, S. (1972) Biochem. Biophys. Acta 268, 221–224
- 3 Muramatsu, M., Shiraishi, S. and Fujii, S. (1972) Biochem. Biophys. Acta 285, 224–234

- 4 Mares-Guia, M. and Shaw, E. (1967) *J. Biol. Chem.* 242, 5782—5788
- 5 Chase, Jr, T. and Shaw, E. (1967) *Biochem. Biophys. Res. Commun.* 29, 508—520
- 6 Bing, D.H. (1969) *Biochemistry* 8, 4503—4510
- 7 Muramatsu, M., Hayakumo, Y., Onishi, T., Sato, T. and Fujii, S. (1969) *J. Biochem. Tokyo*, 65, 329—342
- 8 Sumi, H. and Muramatsu, M. (1974) *Agr. Biol. Chem.* 38, 605—611
- 9 Tamura, Y., Okamura, K., Otsuka, A. and Fujii, S. (1976) *J. Biochem. Tokyo*, 79, 313—319
- 10 Muramatsu, M. and Fujii, S. (1968) *J. Biochem. Tokyo*, 64, 807—814
- 11 Lineweaver, H. and Burk, D. (1934) *J. Am. Chem. Soc.*, 56, 658—666
- 12 Dixon, M. (1953) *Biochem. J.*, 55, 170—171